

Gingival changes during pregnancy: III. Impact of clinical, microbiological, immunological and socio-demographic factors on gingival inflammation

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Abstract

Aims: To identify predictor variables involved in exacerbated gingival inflammation associated with pregnancy.

Material and Methods: In this cohort study, 48 pregnant and 28 non-pregnant women without periodontitis were included. The pregnant women were evaluated in the first, second and third trimester and at 3 months postpartum, whilst the non-pregnant women were evaluated twice, with a 6-month interval. At each visit, clinical [plaque index (PII) and gingival index (GI)], hormonal (salivary progesterone and estradiol), immunological [gingival crevicular fluid interleukin-1 β , interleukin-6, tumour necrosis factor- α (TNF- α) and prostaglandin-E₂] and microbiological (periodontal pathogens culture) evaluations were performed. Statistical analysis was undertaken using exhaustive chi-square automatic interaction detection (exhaustive CHAID) to analyse the predictive value of the independent outcomes to develop pregnancy GI.

Results: PII was the strongest predictor implicated in the GI throughout pregnancy and after delivery. During the second and third trimesters the presence of *Porphyromonas gingivalis* significantly contributed to the worsening of gingival inflammation. When compared with the non-pregnant group, significant differences were found in TNF- α amounts and concentrations and in the third trimester site-specific GI.

Conclusions: Bacterial challenge to the gingival tissues, both quantitatively (PII) and qualitatively (harbouring *P. gingivalis*) appears to affect the level of gingival inflammation observed during pregnancy.

Key words: dental biofilm; IL-6; immunology; microbiology; *Porphyromonas gingivalis*; pregnancy gingivitis; *Prevotella intermedia*; TNF- α

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Conflict of interest and source of funding statement

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Homeostasis in the periodontium involves multiple factors that can modify the clinical expression of plaque-induced gingivitis, including variations in sex hormone production (Armitage 1999). Clinical studies have reported a transient increase in the incidence and severity of

gingival inflammation during pregnancy, unrelated to variations in the amount of plaque present (Silness & Løe 1964, Hugoson 1971, Tilakaratne et al. 2000, Figuero et al. 2010).

During pregnancy, periodontal tissue responses to the biofilm challenge are strengthened, as female sex hormones are necessary but not sufficient to produce gingival changes by themselves and a minimum amount of plaque is required (Arafat 1974, Chaikin 1977). Despite extensive research linking periodontal conditions with sex hormones kinetics, more definitive molecular mechanisms still remain to be determined (Mascarenhas et al. 2003). Models for the hormonal role in the periodontium depend on the understanding of the actions and interactions of hormones with the resident population of cells (Mariotti 1994).

Different aetiological pathways have been proposed in an attempt to explain the increased gingival inflammation observed; however, the results so far have been inconclusive. The most prominent theories to describe pregnancy repercussion include hormone effects on the subgingival biofilm, the immune system, the vasculature and the specific cells of the periodontium. Different studies have dealt with these potential aetiologies, but the response of the periodontium is probably not related to a single mechanism but rather multifactorial in nature (Mariotti 1994). The simultaneous evaluation of the different potential mechanisms may therefore provide a broader understanding of this gingival endocrinopathy.

According to the immune-system theory, immuno-modulative changes developed for foetal tolerance would render periodontal tissues more prone to develop gingival inflammation during pregnancy (O'Neil 1979a, Lopatin et al. 1980, Raber-Durlacher et al. 1991). Interleukin-6 (IL-6), a pleiotropic proinflammatory cytokine, has been suggested to be modulated by hormones, but this proposal is still controversial. Some studies have shown a down-regulation of IL-6 production after sex hormones stimulation, hypothesizing that this would render the gingiva less efficient to bacterial challenge (Cohen-Solal et al. 1993, Lapp et al.

1995, Gornstein et al. 1999, Lapp & Lapp 2005), while other researches reported that the production of IL-6 was significantly enhanced by the stimulation of estradiol and progesterone (Yokoyama et al. 2005). TNF- α is another proinflammatory biomarker, possibly affected by hormonal variations. Oestrogen deficiency has been demonstrated to increase T-cell production of TNF- α (Weitzmann & Pacifici 2006), supporting the concept of type 1-cytokine down-regulation by sex steroids (Szekeres-Bartho 2002, Piccinni 2010, Shiao & Reynolds 2010). On the other hand, animal models have shown that a bacterial challenge with *Porphyromonas gingivalis*, as a localized infection, in pregnant mice triggers an inflammatory response with an increase of TNF- α serum levels (Collins et al. 1994, Lin et al. 2003).

The explanation of an exacerbation of the gingival inflammation, as a consequence of changes in the supra- or subgingival biofilm, is one of the most solidly based hypotheses. A qualitative shift of the subgingival microbiota in a group of non-periodontitis pregnant women that showed an increase in gingival inflammation during pregnancy was recently reported. Significant differences in proportions were found for *Aggregatibacter actinomycetemcomitans*, *P. gingivalis*, *Prevotella intermedia/nigrescens*, *Tannerella forsythia*, *Parvimonas micra*, *Campylobacter rectus* and *Fusobacterium nucleatum* when comparing pregnancy to 3 months postpartum (Carrillo-de-Albornoz et al. 2010). This worsening of the periodontal condition during pregnancy, concomitant with a more pathogenic subgingival microbiota, was in agreement with previous reports (Kornman & Loeche 1980, Raber-Durlacher et al. 1994, Adriaens et al. 2009). This report is the third of a series of articles designed to simultaneously evaluate the role of different potential aetiological pathways on gingival inflammation during pregnancy. In the previous reports, clinical findings, IL-1 β and prostaglandin E2 (PGE2) (Figuero et al. 2010) and the composition of the subgingival microbiota (Carrillo-de-Albornoz et al. 2010) were assessed. The present report aims to evaluate, by means of

a multivariate analysis, the effect of all the analysed factors in the whole series of reports (clinical, socio-demographic, immunological and microbiological) over the gingival inflammation developed in a cohort of pregnant women without periodontitis. Furthermore, the role of additional inflammatory mediators was assessed, specifically whether the higher gingival inflammatory reaction in pregnant women was associated with changes in IL-6 and TNF- α levels in GCF.

Patients and Methods

Study design and patient sample

This was an open cohort prospective study with parallel design, and the methodology, as well as the inclusion and exclusion criteria for subject enrolment have already been reported (Carrillo-de-Albornoz et al. 2010, Figuero et al. 2010). Briefly, two groups (42 pregnant and 20 non-pregnant) of non-smoking, systemically and periodontally healthy women were followed for a 9-month period. Data were gathered on the pregnant women at four visits: at the end of the first trimester (12–14 weeks of pregnancy), second trimester (23–25 weeks of pregnancy) and third trimester (33–36 weeks of pregnancy) and at 3 months postpartum. Data on non-pregnant women were collected at two visits, 6 months apart, matching the first and the third visits of the pregnant group. Hormonal menstrual cycle status was controlled, scheduling visits during the luteal phase (days 17–21). Socio-demographic parameters were registered at the first visit and at each visit clinical, hormonal, immunological and microbiological evaluations were undertaken (Fig. 1).

After all evaluations and sampling procedures at the baseline visit, patients received oral hygiene instructions. To standardize supra-gingival plaque control home care devices, a manual toothbrush (Vitis access[®], Dentaid, Cerdanyola, Spain) and a well-tolerated dentifrice among pregnant women (Colgate Total[®], Palmolive, Piscataway, NJ, USA) (Kraivaphan et al. 2006) were given. No other interventions were performed. At the end of the study, all subjects received a professional prophylaxis.

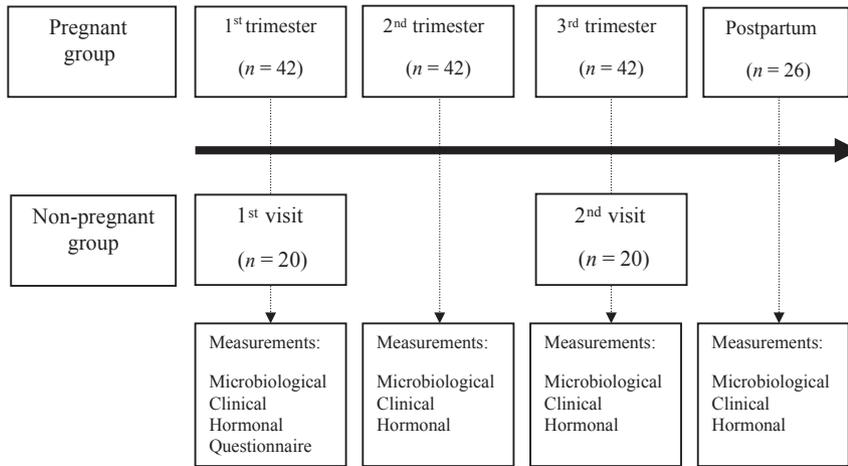


Fig. 1. Flow-chart of follow-up of pregnant and non-pregnant groups. Pregnant women attended four visits and non-pregnant women two visits, corresponding in time to the first and third trimesters of pregnancy. Microbiological, clinical and hormonal assessments were performed at all visits. At the first visit, all women were asked to complete a self-reported questionnaire to assess socio-demographic features and dental care awareness data.

The study design was approved by the Research and Ethics Committee of San Carlos University Hospital (Madrid). Written informed consent was obtained from all participants.

Study variables

At each visit, the following evaluations were carried out, in the order listed below.

- Socio-demographic features. At the first visit, all women were asked to complete a self-reported questionnaire to assess their socioeconomic status (age, education level and profession) and dental care awareness (frequency of tooth brushing, last visit to the dentist and self-evaluation of their oral status).
- Progesterone and estradiol levels in saliva. Unstimulated saliva was collected during 2 min. into a sterile glass tube and stored frozen at -20°C for a maximum time period of 1 year (Morishita et al. 1988, Meulenberg & Hofman 1989). Subsequently, progesterone and estradiol concentrations were calculated by means of a competitive immuno-enzymatic colorimetric method (DIA. METRA S.r.l, Foligno, Italy).
- IL-6, TNF- α , IL-1 β and PGE2 analysis. GCF was collected from

the mesiobuccal sulcus of both upper canines (1.3 and 2.3), using Harco Periopaper (Harco, Irvine, CA, USA) (two samples per patient and per visit). Each strip was measured for fluid volume with a calibrated Periotron 8000[®] (Harco) (Chapple et al. 1999) and placed in a sterile Eppendorf tube. GCF sample from tooth 1.3 was used to measure TNF- α and IL-1 β and that from tooth 2.3 to measure IL-6 and PGE2, using enzyme-linked immunosorbent assays (ELISA) (IL-6, TNF- α , IL-1 β : BLK Diagnostics International, Badalona, Barcelona, Spain; PGE2: DRG Diagnostic, DRG Instruments GmbH, Marburg; Germany). Analyses were performed according to the manufacturer's protocol. Results were calculated using the standard curves created for each assay. Concentrations were corrected for GCF volume and defined as nanograms per millilitre. The total amount of IL-6, TNF- α , IL-1 β and PGE2 was expressed in picograms.

- Plaque index (PII), according to Silness & L oe (1964). This parameter was recorded in all teeth at four sites per tooth (mesial, distal, buccal and lingual) with a CPC-12 periodontal probe (Hu-Friedy, Leinmen, Germany). A site-specific evaluation was obtained from the immuno-

logical sampled sites (sPII), in addition to full-mouth PII recording.

- Microbiological assessment. A pooled subgingival sample was obtained from four sampled sites. At each visit, the sample was obtained from the four inter-proximal sites showing the most marked inflammation per quadrant, excluding those sites in which GCF samples were taken. Samples were obtained with two sterile consecutive #30 paper points per site (Zipperers, United Dental MFRS Inc., West Palm Beach, FL, USA) and transferred to a vial containing 1.5 ml of reduced transport fluid (RTF) (Syed & Loesche 1972). At the laboratory, aliquots of 0.1 ml were plated manually for the detection of *A. actinomycetemcomitans* on the specific medium DentaId-1 (Alsina et al. 2001). Suspected isolates were identified on the basis of colony morphology and positive catalase reaction. Sample dilutions were also plated onto a non-selective blood agar plate (Blood Agar Base II[®], Oxoid, Basingstoke, UK), supplemented with haemine (5 mg/l), menadione (1 mg/l) and 5% sterile horseblood. Suspected colonies were identified by microscopy, confirming their identification by means of Gram staining and cell morphology, aero tolerance, production of catalase and other biochemical reactions (Rapid ID 32A, BioMerieux SA, Le-Balmeles-Grottes, France). Total anaerobic counts were calculated, as well as counts of the detected periodontal pathogens (*P. intermedia/nigrescens*, *P. gingivalis*, *A. actinomycetemcomitans*, *T. forsythia*, *F. nucleatum*, *P. micra*, *Eikenella corrodens*, *C. rectus* and *Capnocytophaga* sp.). In addition to the quantitative microbiological data, the frequency of detection and proportions for each bacterial species were also calculated.
- Gingival index (GI), according to L oe & Silness (1963). GI was recorded at all sites at four sites per tooth (mesial, distal, buccal and lingual) with a CPC-12 periodontal probe (Hu-Friedy, Leinmen, Germany). A site-specific evaluation was obtained from the

immunological sampled sites (sGI), in addition to full-mouth GI recording.

All laboratory analyses (microbiological and immunological) were performed at the Research Laboratory, School of Dentistry, at the University Complutense, Madrid (Spain).

Data management and statistical analysis

Interleukin-6 and TNF- α analysis

A subject level analysis was performed for each study outcome. Biomarker data (IL-6 and TNF- α) were expressed as amounts (pg) and concentrations (ng/ml). Clinical measurements were averaged across subjects from the sites selected for GCF sampling for immunological analysis (sPII and sGI).

Kolmogorov-Smirnov test was applied for each variable to assess the “goodness of fit” to normal distribution. As normality was not achieved rigorously for all the variables at the different time point intervals, non-parametric tests were used. Data were expressed by median and interquartile range. Intra-group differences to evaluate longitudinal variations over time were determined by Friedman’s test. Post hoc comparisons were performed using Bonferroni’s corrections. To determine differences between pregnant and non-pregnant women (inter-group comparison), the Mann–Whitney *U*-test was used.

Exhaustive CHAID algorithm

In order to identify predictors that could explain the increased gingival

inflammation associated to pregnancy, the influence of the different aetiological potential factors evaluated in the present study was analysed in a multivariate analysis. A decision tree through the Chi-square automatic interaction detector (SPSS Exhaustive CHAID) was used, as it offered the capacity to combine categorical and continuous variables. This procedure is a non-parametric analysis based on statistically recursive partitioning algorithms that classify independent variables into predictor values of a dependent outcome. GI was set as the dependent outcome, while 30 independent outcomes were tested: (i) clinical (full mouth PII), (ii) socio-demographic (age, education level, profession, frequency of tooth brushing, frequency of dentist visiting and self-perception of oral health); (iii) immunological (IL-1 β , IL-6, TNF- α , PGE2) and (iv) microbiological variables (total anaerobic bacterial counts and specific counts and percentage of the following periodontal pathogens: *P. intermedia/nigrescens*, *P. gingivalis*, *A. actinomycetemcomitans*, *T. forsythia*, *F. nucleatum*, *P. micra*, *E. corrodens*, *C. rectus* and *Capnocytophaga* sp). Significance values for merging and splitting criteria were adjusted using the Bonferroni method. As inflammatory mediators were site-specific measured, to further explore these outcomes another exhaustive CHAID algorithm analysis was performed, where sGI was set as the dependent outcome and sPII was added to the independent outcomes.

Exhaustive CHAID analysis results are presented in decision trees, which provide a hierarchical

visual depiction of predictor variables interactions. The independent variables that were significantly associated with the dependent outcome are classified in nodes. The variable at the highest level of the tree is determined to have the closest statistical association with the dependent outcome, and is represented as the first node. Subsequent associations are classified in consecutive nodes. Groups were split under the following criteria: tree depth was limited to three levels and the minimum number of cases per node for parent nodes was ten. For child nodes, the minimum number of cases was set at 10% of the sample size.

Statistical significance was established at the 95% confidence level. SPSS for Windows (SPSS Inc. version 18) was used for the data analysis.

Results

The results for all the parameters, except IL-6 and TNF- α , have been previously analysed separately (Carrillo-de-Albornoz et al. 2010, Figuero et al. 2010).

Patient sample

Out of a sample of 60 pregnant women, 48 agreed to participate in the study. Forty-two women (30.15 years, range 20–35) completed the three visits during pregnancy and the postpartum visit was completed by 26 women. In the non-pregnant group, 20 non-pregnant women (24.38 years, range 22–26) completed the study from the 28 women initially enrolled in the study (Fig. 1).

Table 1. Site-specific plaque index (sPII) and gingival index (sGI) scores [median (inter-quartile range)] in GCF sample sites for pregnant and non-pregnant women

	Pregnant group			
	First trimester (<i>n</i> = 42)	Second trimester (<i>n</i> = 42)	Third trimester (<i>n</i> = 42)	Postpartum (<i>n</i> = 26)
sPII	0.50 (0.0–1.0)	0.00 (0.0–0.50)	0.00 (0.0–0.50)	0.50 (0.0–0.50)
sGI	0.75 (0.00–1.50)	1.00* (0.50–2.00)	1.00† (0.50–2.00)	0.25* (0.00–1.00)
	Non-pregnant group			
	Baseline (<i>n</i> = 20)		6 months (<i>n</i> = 20)	
sPII	0.00 (0.00–0.50)		0.00 (0.00–0.50)	
s-GI	0.25 (0.00–1.00)		0.50† (0.00–1.00)	

Intragroup comparison: Friedman’s test with Bonferroni correction (**p* < 0.05); intergroup comparison: Mann–Whitney *U*-test (†*p* < 0.05).

Table 2. IL-6 levels (amount and concentration) in gingival crevicular fluid of pregnant and non-pregnant women

	Pregnant group			
	First trimester (n = 42)	Second trimester (n = 42)	Third trimester (n = 42)	Postpartum (n = 26)
Amount (pg)	0.08 (0.01–0.14)	0.12 (0.02–0.20)	0.16 (0.03–0.27)	0.04* (0.0–0.22)
Concentration (ng/ml)	0.24 (0.02–0.62)	0.27 (0.06–0.81)	0.66 (0.16–1.02)	0.11* (0.0–0.56)
	Non-pregnant group			
	Baseline (n = 20)		6 months (n = 20)	
Amount (pg)	0.01 (0.0–0.12)		0.06 (0.0–0.22)	
Concentration (ng/ml)	0.05 (0.0–0.3)		0.26 (0.0–1.39)	

IL-6 levels are expressed as median (inter-quartile range). Intragroup comparison: Friedman's test with Bonferroni correction (* $p < 0.05$); intergroup comparison: Mann–Whitney U -test.

Clinical parameters

Site-specific plaque and gingivitis levels were analysed from the GCF isolated sampled teeth. sPII showed a slight decrease during pregnancy and a tendency to increase after delivery, but changes were not significant. sGI increased significantly from the first trimester to the second trimester ($p = 0.02$), then maintained high levels at the third trimester, and decreased postpartum ($p < 0.001$) (Table 1).

Interleukin-6 and TNF- α levels in GCF

Table 2 shows the longitudinal changes of IL-6 during pregnancy. Both IL-6 amounts and concentrations showed a gradual increase throughout pregnancy and decreased after delivery, concomitant with the decrease in the sGI. Changes were statistically significant for the reduction from the third trimester to postpartum visit ($p < 0.001$).

TNF- α behaviour in CGF throughout pregnancy is depicted in Table 3. Amounts and concentrations of this inflammatory mediator

decreased progressively from the first to the third trimester. At the postpartum visit, the concentration of the biomarker increased. Changes were significant for the reduction in the absolute quantity from the second to the third trimester ($p = 0.03$).

Comparison between pregnant and non-pregnant women

The non-pregnant group showed minor changes in the studied outcomes between the two visits (Tables 1–3). When compared with the pregnant group, significant differences were found in the sGI between the third trimester (pregnant group) and the 6-month visit for the non-pregnant group (Table 1). TNF- α amounts and concentrations were significantly higher in the pregnant group, both at the first and third trimester comparison.

Gingival index CHAID decision tree analysis

Decision trees were built up for each trimester, analysing the impact of

the 30 independent outcomes evaluated: (i) clinical (full mouth PII), (ii) socio-demographic (age, education level, profession, frequency of tooth brushing, frequency of dentist visiting and self-perception of oral health); (iii) immunological (IL-1 β , IL-6, TNF- α , PGE2) and (iv) microbiological variables (total anaerobic bacterial counts and specific counts and percentage of the following periodontal pathogens: *P. intermedia/nigrescens*, *P. gingivalis*, *A. actinomycetemcomitans*, *T. forsythia*, *F. nucleatum*, *P. micra*, *E. corrodens*, *C. rectus* and *Capnocytophaga* sp.), over the gingival inflammation (GI).

First trimester

PII was the only factor significantly involved with first term GI (Fig. 2). None of the other outcomes influenced GI at this stage.

Four nodes were obtained depending on PII values. At a given PII minor or equal to 0.35, the expected GI at the first term was 0.49 (node 1). In the second node, where the PII raised (0.35–0.51), the

Table 3. TNF- α levels (amount and concentration) in gingival crevicular fluid of pregnant and non-pregnant women

	Pregnant group			
	First trimester (n = 42)	Second trimester (n = 42)	Third trimester (n = 42)	Postpartum (n = 26)
Amount (pg)	3.99 [†] (1.70–9.33)	2.37 (1.07–11.44)	1.30* [†] (0.35–9.69)	1.31 (0.84–7.31)
Concentration (ng/ml)	9.07 [†] (5.77–30.68)	8.84 (4.01–48.10)	4.84 [†] (0.99–24.25)	9.56 (1.75–26.98)
	Non-pregnant group			
	Baseline (n = 20)		6 months (n = 20)	
Amount (pg)	0.45 [†] (0.09–0.59)		0.40 [†] (0.09–0.82)	
Concentration (ng/ml)	1.55 [†] (0.21–2.94)		1.86 [†] (0.74–2.92)	

TNF- α levels are expressed as median (inter-quartile range). Intragroup comparison: Friedman's test with Bonferroni correction (* $p < 0.05$); intergroup comparison: Mann–Whitney U -test ([†] $p < 0.05$).

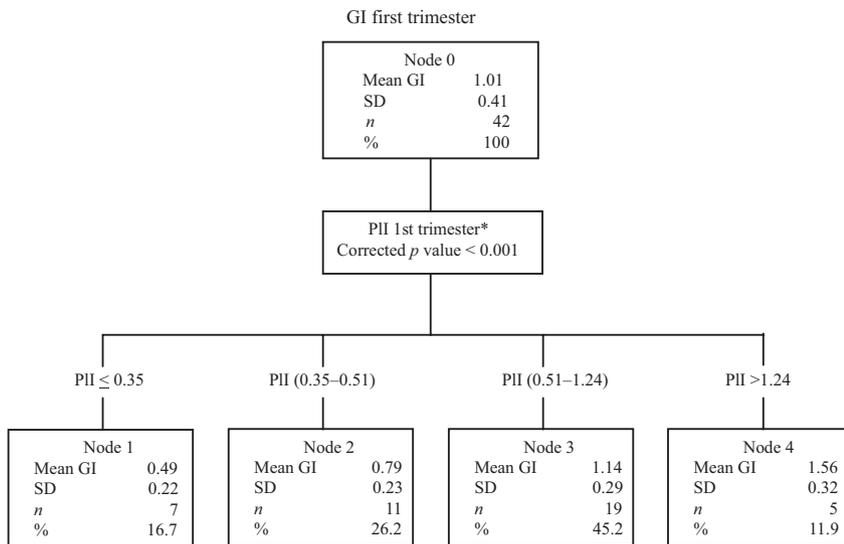


Fig. 2. Decision tree for gingival index (GI) in the first trimester (Exhaustive CHAID algorithm). PII, Plaque index; SD, Standard deviation. * $p < 0.05$.

GI increased to 0.79. The same pattern was observed at the third and fourth nodes, where PII progressively increased and so the GI did (1.14 and 1.56, respectively).

Second trimester

The predictors significantly associated with GI were PII and counts of *P. gingivalis*. PII was the main outcome implicated in the gingival inflammation during second trimester. Three groups were obtained depending on different second trimester PII range scores ($p < 0.001$), with GI of 0.72, 1.23 and 1.85 for nodes 1, 2 and 3, respectively.

Node 2 was the most prevalent, including 61.9% of the cases and it was significantly associated with counts of *P. gingivalis*. Counts of *P. gingivalis* of less than 6600 colonies were associated with a GI of 1.09, while for counts exceeding 6600 colonies, the GI increased to 1.48 (Fig. 3).

Third trimester

Outcomes followed the same pattern as described for the second trimester. PII presented on the second trimester was the main involved variable ($p < 0.001$). Three filial nodes were derived, with different GI scores depending on the PII values. Node 2, again the most prevalent, presented a GI of 1.22 and was also associated with the counts of *P. gingivalis*. Absence of *P. gingivalis* was con-

comitant with less inflammation, while the presence of the pathogen was associated with an increased GI (Fig. 4).

Postpartum

PII at first trimester during pregnancy was the main outcome responsible for GI observed after delivery. If pregnant women presented a PII of less than 0.43 in the first trimester, the GI expected after delivery was 0.61 (node 1). On the other hand, for those women with a first trimester PII value higher than 0.43, GI increased to 1.14 after delivery (node 2). The gingival inflammation also depended on the counts of *P. micra* and PII during the second term (Fig. 5).

Site-specific GI CHAID decision tree analysis

Inflammatory mediators (IL-1 β , IL-6, TNF- α , PGE2) were not significantly involved in the corresponding site-specific GI developed during pregnancy (data not shown).

Discussion

The present study aimed to evaluate the overall effect of different potential factors over gingival inflammation during pregnancy. A multivariate analysis was used to determine the impact of the clinical, socio-demographic, immunological

and microbiological factors over the associated GI. This analysis revealed that PII presented during pregnancy was the main implicated outcome in the GI developed throughout pregnancy and after delivery. During the second and third trimesters, *P. gingivalis* was implicated in the worsening of the clinical condition. This implies that both PII and harbouring *P. gingivalis* could modify gingival inflammation during pregnancy.

Prevalence, extent and severity of pregnancy gingivitis vary considerably among different studies. Methodological heterogeneity may, at least in part, explain differences in the obtained results. Cross-sectional (Løe & Silness 1963, Silness & Løe 1964, Katz et al. 1969, Adams et al. 1974, Arafat 1974, Samant et al. 1976, Conde Vidal et al. 1981, Zaki et al. 1984, Jonsson et al. 1988, Miyazaki et al. 1991, Muramatsu & Takaesu 1994, Kraivaphan et al. 2006, 2007, Acharya & Bhat 2009), in comparison to longitudinal studies (Cohen et al. 1969, Hugoson 1971, Chaikin 1977, O'Neil 1979b, Machuca et al. 1999, Tilakaratne et al. 2000, Yalcin et al. 2002, Gürsoy et al. 2008), hamper the analysis of the relationship between pregnancy and the exacerbation of gingival inflammation (Stroup et al. 2000, von Elm et al. 2008). Other factors that vary within different research groups may have affected the wide range in the results obtained, including the use of different clinical indices, study designs, measurement equipments and the control of confounding factors.

To evaluate the exposure of gingival tissues to pregnancy, the present cohort study was designed in an attempt to standardize methodology and overcoming previous reported limitations, thus results were reported following the STROBE statement (von Elm et al. 2008). As detailed in the previous reports (Carrillo-de-Albornoz et al. 2010, Figuero et al. 2010), it is important to highlight the weaknesses of the study, including the high incidence of dropouts (specially after delivery) and the lack of homogeneity between the groups in terms of demographic characteristics and initial clinical status, which may limit

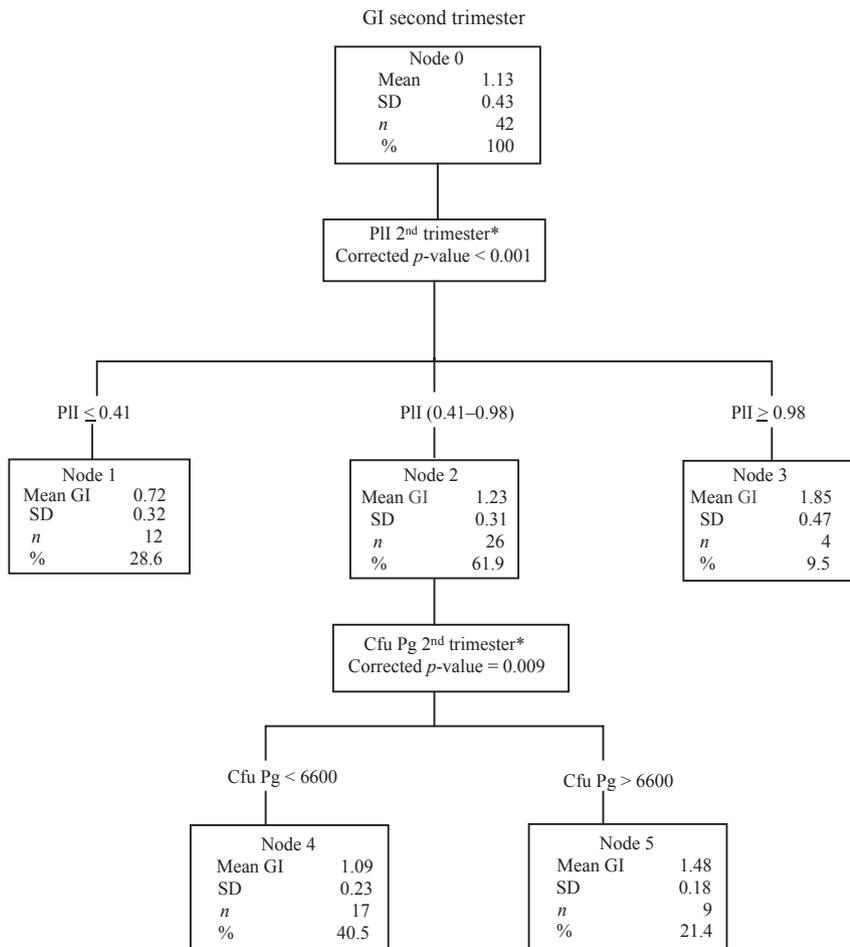


Fig. 3. Decision tree for gingival index (GI) in the second trimester (Exhaustive CHAID algorithm). PII, Plaque index; SD, Standard deviation; Cfu Pg, Colony forming units of *P. gingivalis*. * $p < 0.05$.

the comparison between pregnant and non pregnant groups.

To standardize supragingival plaque control home care, a well-tolerated dentifrice among pregnant women was selected (Kraivaphan et al. 2006). Within the context of the present study, it is important to remark relevant properties of triclosan/copolymer formulation, including inhibition of periodontal pathogens such as *A. actinomycetemcomitans*, *E. corrodens* and *F. nucleatum* (Haraszthy et al. 2010). In vitro studies also support an anti-inflammatory activity, based upon suppression of acute and chronic mediators of inflammation (Gaffar et al. 1995, Barros et al. 2010).

Gingival index exhaustive CHAID analysis

The predictive potential of data mining algorithms to medical research is becoming an increasingly used statis-

tical tool, particularly in recent years. Decision trees are being widely applied due to its capacity for increasing efficacy and managing study quality in medicine by the implementation of specified standards into a systematic, logical, evidence-based and rational concept (Greco et al. 2010, Khalil et al. 2010). In gingival inflammatory changes associated with pregnancy, no predictive models have been thus far reported.

The exhaustive CHAID analysis revealed PII to be the main predictor outcome implicated in the GI of pregnant women during all trimesters. Pregnancy gingivitis is described as a gingival inflammatory condition initiated by plaque and exacerbated by sex steroid hormones (Mariotti 1999). A minimum amount of plaque is required, as pregnant women with excellent plaque control do not develop it or reduce its incidence to

0.03% (Arafat 1974, Chaikin 1977). Several studies have reported an increase in gingival inflammation during pregnancy without associated changes in plaque levels (Silness & Løe 1964, Cohen et al. 1969, 1971, Hugoson 1971, O'Neil 1979a, Zaki et al. 1984, Tilakaratne et al. 2000). This renders plaque as a necessary factor, but plays down its importance over the characteristic gingival inflammation associated to pregnancy.

Previous attempts have been made to relate the PII to the GI. Silness & Løe (1964) found a 0.73 GI/PII correlation coefficient during pregnancy and 0.99 at postpartum visit. This diminishes the role of the plaque component and suggests that other factors are implicated in gingival inflammation. In agreement with the results of the present study, Kinby et al. (1996) found a significant increased reactivity to plaque during pregnancy after calculating the sites with gingivitis to sites with bacterial plaque (G/P-ratio). In this study it was observed that, without intervention, the plaque level presented during the first and second trimesters of gestation modulates the increased gingival inflammation during pregnancy. This highlights the relationship between the extent of plaque deposits and the severity of gingivitis, as GI during pregnancy could be estimated on the amount of plaque present.

The gingival inflammation present during the second and third trimesters of pregnancy could be explained in 61.9% of the cases by qualitative changes in the microbial composition of the subgingival biofilm. This group of pregnant women cover the filial nodes generated from the parent node with intermediate plaque level (Node 2; PII 0.41–0.98). *P. gingivalis* counts were the principal factor responsible for the splitting into different GI filial nodes. Absence of *P. gingivalis* was associated with low GI scores, while the presence of the pathogen was associated with a worsening of the periodontal condition. This means, at least in part, that harbouring *P. gingivalis* at different counts and proportions during pregnancy favours the development of more severe forms of gingivitis and explains differences in the clinical

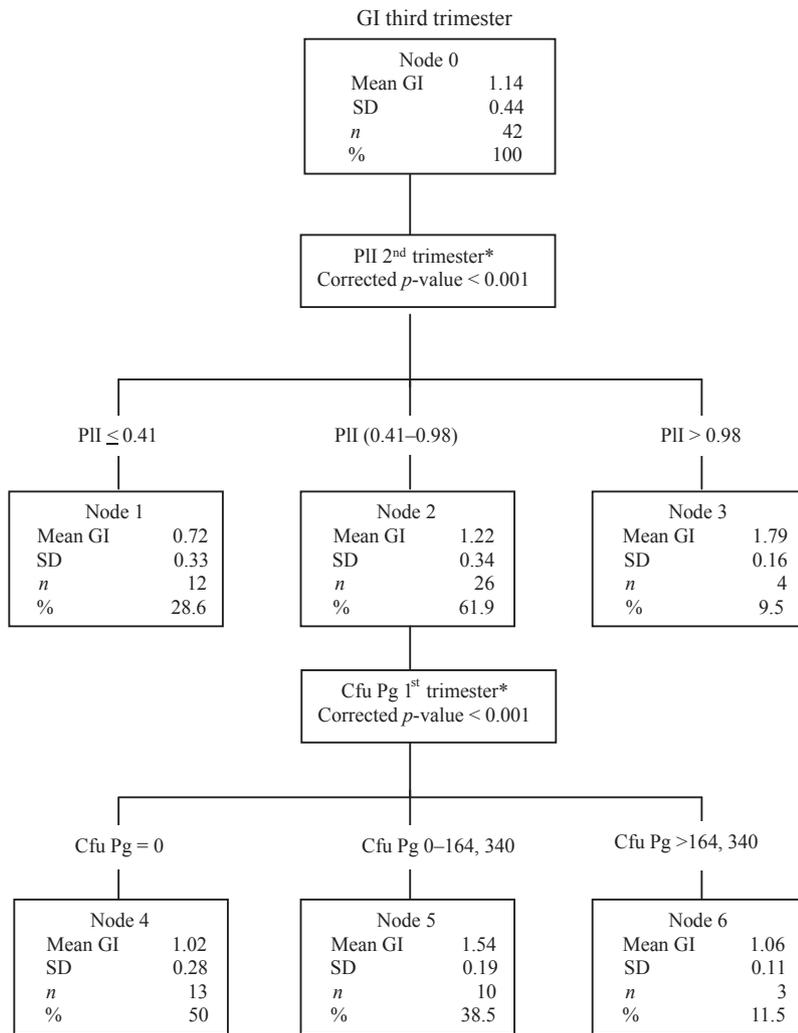


Fig. 4. Decision tree for gingival index (GI) in the third trimester (Exhaustive CHAID algorithm). PII, Plaque index; SD, Standard deviation; Cfu Pg, Colony forming units of *P. gingivalis*. **p* < 0.05.

presentation amongst the pregnant women. The frequency of detection of *P. gingivalis* in the present study was relatively constant throughout pregnancy (35.7–40.5%) and proportions of microbiota in positive subjects ranged from 11.2% to 20.1% (Carrillo-de-Albornoz et al. 2010). These results corroborate the findings of previous reports of *P. gingivalis* detection in Spanish population with gingivitis (Lau et al. 2004), considering that Spain presents a high prevalence of these bacteria in comparison to other European countries, the Netherlands in particular, using identical bacteriological methods (Sanz et al. 2000). Geographical differences in the composition of the subgingival

biofilm have been observed, although to date there are insufficient data to explain the basis of these differences (Marsh & Devine 2011, Sanz & van Winkelhoff 2011). In periodontally healthy patients the prevalence of *P. gingivalis* has been reported worldwide, being found in 25% in a US population (Griffen et al. 1998), 23.1% in Taiwanese subjects (Yang et al. 2004) and 22.1% in a Chinese adult population (Zhao et al. 2007).

P. gingivalis has repeatedly demonstrated a strong association with disease (Kebuschull & Papapanou 2011). Bacterial species aetiologically related to periodontitis, including *A. actinomycetemcomitans*, *P. gingivalis*, *T. forsythia* and *T. denticola* (aetio-

logic burden group), have been associated with increased bleeding on probing prevalence in sites with PD ≤ 3 mm (Demmer et al. 2008). In patients with periodontitis, *P. gingivalis* has been reported to be more frequently detected in destructive forms (Haffajee & Socransky 1994, Takeuchi et al. 2001, van Winkelhoff et al. 2002) and in subjects exhibiting periodontal disease progression (Grossi et al. 1995). In successfully treated patients it appears to be significantly reduced and it is commonly encountered in sites that exhibit recurrence and progression (Mombelli et al. 2000, Fujise et al. 2002). Two postpartum parenteral nodes were formed depending on the first trimester PII. The higher PII was related to a worsening of the clinical condition. The Node 2 group presented with an increased gingival inflammation (GI 1.14) that was associated with the presence and counts of *P. micra* and additionally with the second trimester PII. Counts of *P. micra* exceeding 7260 colonies were concomitant with a reduction of the GI, but it is important to highlight that no definitive conclusions can be drawn, as only three cases were included in this group.

Several authors have proposed the increase in *P. intermedia/nigrescens* counts during pregnancy as an aetiological factor in pregnancy gingivitis (Kornman & Loesche 1980, Jensen et al. 1981, Raber-Durlacher et al. 1994). Gürsoy et al. (2009) reported that the vast majority of isolates of *P. intermedia sensu lato* during pregnancy proved to be *P. nigrescens* (95.3%). Our research group observed a significant increase in the gingival inflammation in pregnant *P. intermedia/nigrescens*-positive women, but considering that this was concomitant with an increase in plaque levels (Carrillo-de-Albornoz et al. 2010). In the present study, *P. intermedia/nigrescens* was not a predictor outcome involved in the GI developed during pregnancy, what is in agreement with other reports that found no microbiological differences in the mentioned pathogens (Jonsson et al. 1988, Adriaens et al. 2009).

From an aetiopathogenic point of view, the influence of bacterial challenge over gingival inflammation during pregnancy can be direct or

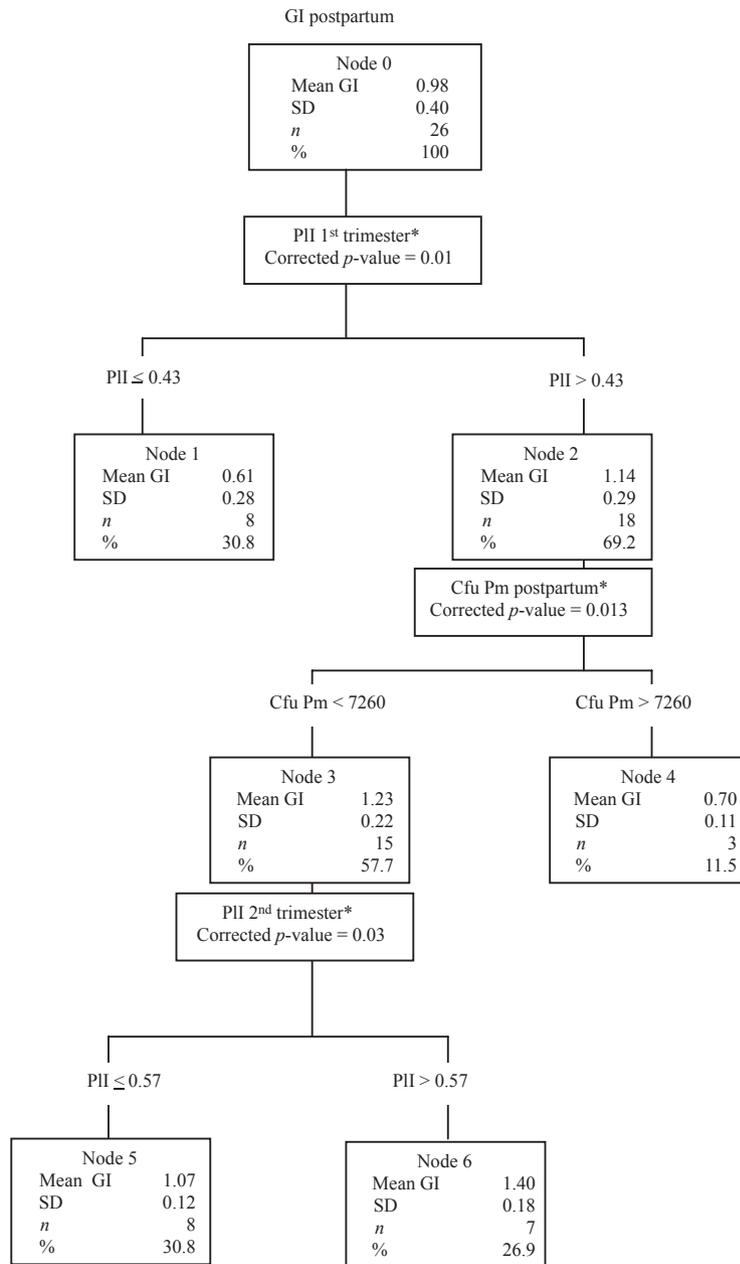


Fig. 5. Decision tree for gingival index (GI) at postpartum visit (Exhaustive CHAID algorithm). PII, Plaque index; SD, Standard deviation; Cfu Pm, Colony forming units of *P. micra*. * $p < 0.05$.

indirectly explained. According to the direct pathway, the increase in hormone levels would promote the overgrowth of specific bacteria that are responsible for the increased gingival inflammation (Kornman & Loesche 1982). However, an indirect pathway can not be discarded, assuming that the presence of the pathogen would be the consequence and not the cause of the condition, as the greater exposure to sex steroid

hormones would transform the gingiva into a more susceptible environment due to greater gingival probing depths (Miyazaki et al. 1991), a higher gingival crevicular flow rate (Lindhe & Branemark 1968), a lower degree of keratinization (Mariotti 1994) and reduced immunoresponsiveness.

None of the other microbiological or immunological factors evaluated in the present model were signifi-

cantly implicated in the gingival inflammation developed during pregnancy. Further research is therefore warranted, analysing other aetiological variables and increasing the sample size to corroborate the present results.

Interleukin-6 and TNF- α levels

Sex steroids have been suggested to contribute to observed maternal immune-modulation during pregnancy (Mellor & Munn 2000, Szekeres-Bartho 2002, Shiao & Reynolds 2010). Immunologic tolerance during successful pregnancy seems to be associated with an increase in humoral immunity (type-2 cytokine production) and a decrease in cell-mediated immune response (type-1 cytokine production) (Wegmann et al. 1993). Repercussion of sex steroids over periodontal conditions is furthermore recognized in situations of oestrogen deficiency, as seen in post-menopausal women (Haas et al. 2009).

Different in vitro studies have shown a potential inhibitory effect of sex hormones over the IL-6 secretion (Cohen-Solal et al. 1993, Lapp et al. 1995, Gornstein et al. 1999, Lapp & Lapp 2005). Telleria et al. (1998) corroborated in an animal study the down-regulation of IL-6 production, although the expression of IL-6 mRNA increased after an in vivo injection of bacterial lipopolysaccharide. This contrasts with other studies in which IL-6 production increased following estradiol or progesterone stimulation, at concentrations comparable to those presented in the plasma of pregnant women. This finding suggests that large amounts of female sex hormones in pregnant women may directly stimulate the production of this cytokine by gingival fibroblasts (Yokoyama et al. 2005).

In the present study, IL-6 (amounts and concentrations) progressively increased during pregnancy and was significantly reduced after delivery, concomitant with the fall in hormone production. This is in agreement with previous human studies that correlate local levels of IL-6 with the periodontal status at different life stages. Furthermore, chronic gingivitis has been associated with an increase in IL-6 levels in

GCF when compared to periodontally healthy subjects (Offenbacher et al. 2007). Becerik et al. (2010) reported that GCF levels of IL-6 were significantly higher in a group of women with gingivitis (bleeding on probing – BoP – in more than 50% of the probing sites) compared to a group of periodontally healthy women (BoP < 10%), although no differences were found at the different menstrual cycle stages in both groups. Offenbacher et al. (2006) observed that pregnant women with untreated periodontitis, after supra-gingival debridement and a recommendation (without instructions) to use a manual toothbrush, suffered a significant increase in IL-1 β in GCF and a 2.1-fold increase in IL-6 in GCF. Conversely, the intervention group (treated with scaling and root planing, tooth polishing and instructed to use a sonic powered toothbrush) resulted in significant improvements in the clinical status. There were also significant decreases in the levels of serum IL-6 soluble receptor (IL-6sr), IL-1 β in GCF and in levels of *P. intermedia/nigrescens*. Other studies have not found significant reductions in IL-6 levels after non surgical periodontal treatment delivered before 21 weeks of gestation, considering that the biomarker measurement was made in serum (Michalowicz et al. 2009).

TNF- α underwent a down-regulation throughout pregnancy. It was statistically significantly reduced at the third trimester, and increased after delivery. This is in agreement with previous reports that support an inhibitory effect of sex steroid hormones over TNF- α (Cohen-Solal et al. 1993, Weitzmann & Pacifici 2006, Luo et al. 2010).

Anti-inflammatory cytokines may play a key role in the survival to term of the foetal allograft, by counteracting deleterious inflammatory Th-1 cytokines (Szekeres-Bartho 2002). Animal studies in mice have reported that subcutaneous infection of *P. gingivalis*, with the chamber model, increases maternal TNF- α and enhances adverse pregnancy outcomes (Collins et al. 1994, Lin et al. 2003). In clinical studies, limited data are available analysing local TNF- α variations on fluctuations of the sex hormone levels. In non-pregnant women with low standardized

PII scores, Baser et al. (2009) did not observed changes in TNF- α levels in the GCF throughout menstrual cycle stages. In serum, Hasegawa et al. (2003) obtained an increased TNF- α level concomitant with significantly higher mean probing depths, while Michalowicz et al. (2009) did not observed serum changes in any evaluated biomarker (C-reactive protein, PGE2, matrix metalloproteinase-9, fibrinogen, endotoxin, IL-1 β , IL-6, IL-8 and TNF- α) after non surgical periodontal treatment in pregnant women with periodontitis.

Conclusions

Within the limitations of the present study, it can be concluded that the amount of plaque deposits and the presence of *P. gingivalis* above culture threshold were the main implicated factors in the gingival inflammation developed throughout pregnancy, suggesting that these quantitative and qualitative differences in the dental biofilm are able to trigger the inflammatory condition. However, an indirect aetiopathogenic pathway of these factors should also be considered.

Minor significant differences were detected for GCF IL-6 and TNF- α , but these pro-inflammatory biomarkers could not be associated with the gingival inflammatory condition present during pregnancy. Further studies are needed to clarify mechanisms of pregnancy-related gingival inflammation.

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References

- Acharya, S. & Bhat, P. V. (2009) Oral-health-related quality of life during pregnancy. *Journal of Public Health Dentistry* **69**, 74–77.
- Adams, D., Carney, J. S. & Dicks, D. A. (1974) Pregnancy gingivitis: a survey of 100 antenatal patients. *Journal of Dentistry* **2**, 106–110.
- Adriaens, L. M., Alessandri, R., Spörri, S., Lang, N. P. & Persson, G. R. (2009) Does pregnancy have an impact on the subgingival microbiota?. *Journal of Periodontology* **80**, 72–81.
- Alsina, M., Olle, E. & Frias, J. (2001) Improved, low-cost selective culture medium for *Actinobacillus actinomycetemcomitans*. *Journal of Clinical Microbiology* **39**, 509–513.
- Arafat, A. H. (1974) Periodontal status during pregnancy. *Journal of Periodontology* **45**, 641–643.
- Armitage, G. C. (1999) Development of a classification system for periodontal diseases and conditions. *Annals of Periodontology* **4**, 1–6.
- Barros, S. P., Wirojchanasak, S., Barrow, D. A., Panagakos, F., Devizio, W. & Offenbacher, S. (2010) Triclosan inhibition of acute and chronic inflammatory gene pathways. *Journal of Periodontology* **37**, 412–418.
- Baser, U., Cekici, A., Tanrikulu-Kucuk, S., Kantarci, A., Ademoglu, E. & Yalcin, F. (2009) Gingival inflammation and interleukin-1 beta and tumor necrosis factor-alpha levels in gingival crevicular fluid during the menstrual cycle. *Journal of Periodontology* **80**, 1983–1990.
- Becerik, S., Ozçaka, O., Nalbantsoy, A., Atilla, G., Celec, P., Behuliak, M. & Emingil, G. (2010) Effects of menstrual cycle on periodontal health and gingival crevicular fluid markers. *Journal of Periodontology* **81**, 673–681.
- Carrillo-de-Albornoz, A., Figuero, E., Herrera, D. & Bascones-Martínez, A. (2010) Gingival changes during pregnancy: II. Influence of hormonal variations on the subgingival biofilm. *Journal of Clinical Periodontology* **37**, 230–240.
- Chaikin, B. S. (1977) Incidence of gingivitis in pregnancy. *Quintessence International Dental Digest* **8**, 81–89.
- Chapple, I. L., Landini, G., Griffiths, G. S., Patel, N. C. & Ward, R. S. (1999) Calibration of the Periotron 8000 and 6000 by polynomial regression. *Journal of Periodontal Research* **34**, 79–86.
- Cohen, D. W., Friedman, L., Shapiro, J. & Kyle, G. C. (1969) A longitudinal investigation of the periodontal changes during pregnancy. *Journal of Periodontology* **40**, 563–570.
- Cohen, D. W., Shapiro, J., Friedman, L., Kyle, G. C. & Franklin, S. (1971) A longitudinal investigation of the periodontal changes during pregnancy and fifteen months post-partum. II. *Journal of Periodontology* **42**, 653–657.
- Cohen-Solal, M. E., Graulet, A. M., Denne, M. A., Gueris, J., Baylink, D. & de Vernejoul, M. C. (1993) Peripheral monocyte culture supernatants of menopausal women can induce bone resorption: involvement of cytokines. *The Journal of Clinical Endocrinology and Metabolism* **77**, 1648–1653.
- Collins, J. G., Windley, H. W. III, Arnold, R. R. & Offenbacher, S. (1994) Effects of a *Porphyromonas gingivalis* infection on inflammatory mediator response and pregnancy outcome in hamsters. *Infection and Immunity* **62**, 4356–4361.
- Conde Vidal, J. M., Ingles Castello, M. & Roldán González, P. (1981) Periodontal disease during pregnancy. *Revista Española de Estomatología* **29**, 179–190.

- Demmer, R. T., Papapanou, P. N., Jacobs, D. R. Jr & Desvarieux, M. (2008) Bleeding on probing differentially relates to bacterial profiles: the Oral Infections and Vascular Disease Epidemiology Study. *Journal of Clinical Periodontology* **35**, 479–486.
- von Elm, E., Altman, D. G., Egger, M., Pocock, S. J., Gøtzsche, P. C. & Vandenbroucke, J. P. (2008) The Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. *Journal of Clinical Epidemiology* **61**, 344–349.
- Figuero, E., Carrillo-de-Albornoz, A., Herrera, D. & Bascones-Martínez, A. (2010) Gingival changes during pregnancy (I). Influence of hormonal variations on clinical and immunological parameters. *Journal of Clinical Periodontology* **37**, 220–229.
- Fujise, O., Hamachi, T., Inoue, K., Miura, M. & Maeda, K. (2002) Microbiological markers for prediction and assessment of treatment outcome following non-surgical periodontal therapy. *Journal of Periodontology* **73**, 1253–1259.
- Gaffar, A., Scherl, D., Afflitto, J. & Coleman, E. J. (1995) The effect of triclosan on mediators of gingival inflammation. *Journal of Clinical Periodontology* **22**, 480–484.
- Gornstein, R. A., Lapp, C. A., Bustos-Valdes, S. M. & Zamorano, P. (1999) Androgens modulate interleukin-6 production by gingival fibroblasts in vitro. *Journal of Periodontology* **70**, 604–609.
- Greco, R., Papalia, T., Lofaro, D., Maestriperi, S., Mancuso, D. & Bonofiglio, R. (2010) Decisional trees in renal transplant follow-up. *Transplantation proceedings* **42**, 1134–1136.
- Griffen, A. L., Becker, M. R., Lyons, S. R., Moeschberger, M. L. & Leys, E. J. (1998) Prevalence of *Porphyromonas gingivalis* and periodontal health status. *Journal of Clinical Microbiology* **36**, 3239–3242.
- Grossi, S. G., Genco, R. J., Machtei, E. E., Ho, A. W., Koch, G., Dunford, R., Zambon, J. J. & Hausmann, E. (1995) Assessment of risk for periodontal disease. II. Risk indicators for alveolar bone loss. *Journal of Periodontology* **66**, 23–29.
- Gürsoy, M., Haraldsson, G., Hyvönen, M., Sorsa, T., Pajukanta, R. & Könönen, E. (2009) Does the frequency of *Prevotella intermedia* increase during pregnancy? *Oral Microbiology and Immunology* **24**, 299–303.
- Gürsoy, M., Pajukanta, R., Sorsa, T. & Kononen, E. (2008) Clinical changes in periodontium during pregnancy and post-partum. *Journal of Clinical Periodontology* **35**, 576–583.
- Haas, A. N., Rösing, C. K., Oppermann, R. V., Albandar, J. M. & Susin, C. (2009) Association among menopause, hormone replacement therapy, and periodontal attachment loss in southern Brazilian women. *Journal of Periodontology* **80**, 1380–1387.
- Haffajee, A. D. & Socransky, S. S. (1994) Microbial etiological agents of destructive periodontal diseases. *Periodontology 2000* **5**, 78–111.
- Haraszthy, V. I., Zambon, J. J. & Sreenivasan, P. K. (2010) Evaluation of the antimicrobial activity of dentifrices on human oral bacteria. *The Journal of Clinical Dentistry* **21**, 96–100.
- Hasegawa, K., Furuichi, Y., Shimotsu, A., Nakamura, M., Yoshinaga, M., Kamitomo, M., Hatae, M., Maruyama, I. & Izumi, Y. (2003) Associations between systemic status, periodontal status, serum cytokine levels, and delivery outcomes in pregnant women with a diagnosis of threatened premature labor. *Journal of Periodontology* **74**, 1764–1770.
- Hugoson, A. (1971) Gingivitis in pregnant women. A longitudinal clinical study. *Odontologisk Revy* **22**, 65–84.
- Jensen, J., Liljemark, W. & Bloomquist, C. (1981) The effect of female sex hormones on subgingival plaque. *Journal of Periodontology* **52**, 599–602.
- Jonsson, R., Howland, B. E. & Bowden, G. H. (1988) Relationships between periodontal health, salivary steroids, and *Bacteroides intermedius* in males, pregnant and non-pregnant women. *Journal of Dental Research* **67**, 1062–1069.
- Katz, A., Shapiro, S., Gedalia, I. & Zukerman, H. (1969) Periodontal condition and blood citrate level in pregnant women. *Journal of Dental Research* **48**, 140–143.
- Kebschull, M. & Papapanou, P. N. (2011) Periodontal microbial complexes associated with specific cell and tissue responses. *Journal of Clinical Periodontology* **38**(Suppl. 11), 17–27.
- Khalil, P. N., Kleespies, A., Angele, M. K., Thasler, W. E., Siebeck, M., Bruns, C. J., Mutschler, W. & Kanz, K. G. (2010) The formal requirements of algorithms and their implications in clinical medicine and quality management. *Langenbeck's Archives of Surgery* **1**, 31–40.
- Kinby, B., Matsson, L. & Astedt, B. (1996) Aggravation of gingival inflammatory symptoms during pregnancy associated with the concentration of plasminogen activator inhibitor type 2 (PAI-2) in gingival fluid. *Journal of Periodontal Research* **31**, 271–277.
- Kornman, K. S. & Loesche, W. J. (1980) The subgingival microbial flora during pregnancy. *Journal of Periodontal Research* **15**, 111–122.
- Kornman, K. S. & Loesche, W. J. (1982) Effects of estradiol and progesterone on *Bacteroides melaninogenicus* and *Bacteroides gingivalis*. *Infection and Immunity* **35**, 256–263.
- Kraivaphan, P., Amornchat, C. & Triratana, T. (2007) Effects of a triclosan dentifrice on plaque formation, gingivitis and gingival bleeding in pregnant women: five-month clinical results. *The Southeast Asian Journal of Tropical Medicine and Public Health* **38**, 594–597.
- Kraivaphan, P., Amornchat, C., Triratana, T. & Leethochawalit, U. (2006) Clinical effect of a triclosan containing dentifrice on gingivitis during pregnancy and post-partum. *The Southeast Asian Journal of Tropical Medicine and Public Health* **37**, 820–825.
- Lapp, C. A. & Lapp, D. F. (2005) Analysis of interleukin-activated human gingival fibroblasts: modulation of chemokine responses by female hormones. *Journal of Periodontology* **76**, 803–812.
- Lapp, C. A., Thomas, M. E. & Lewis, J. B. (1995) Modulation by progesterone of interleukin-6 production by gingival fibroblasts. *Journal of Periodontology* **66**, 279–284.
- Lau, L., Sanz, M., Herrera, D., Morillo, J. M., Martín, C. & Silva, A. (2004) Quantitative real-time polymerase chain reaction versus culture: a comparison between two methods for the detection and quantification of *Actinobacillus actinomycetemcomitans*, *Porphyromonas gingivalis* and *Tannerella forsythensis* in subgingival plaque samples. *Journal of Clinical Periodontology* **31**, 1061–1069.
- Lin, D., Smith, M. A., Champagne, C., Elter, J., Beck, J. & Offenbacher, S. (2003) *Porphyromonas gingivalis* infection during pregnancy increases maternal tumor necrosis factor alpha, suppresses maternal interleukin-10, and enhances fetal growth restriction and resorption in mice. *Infection and Immunity* **71**, 5156–5162.
- Lindhe, J. & Branemark, P. I. (1968a) Experimental studies on the etiology of pregnancy gingivitis. *Periodontal abstracts* **16**, 50–51.
- Lindhe, J. & Branemark, P. I. (1968b) The effects of sex hormones on vascularization of granulation tissue. *Journal of Periodontal Research* **3**, 6–11.
- Löe, H. & Silness, J. (1963) Periodontal disease in pregnancy. I. Prevalence and severity. *Acta Odontologica Scandinavica* **21**, 533–551.
- Lopatin, D. E., Kornman, K. S. & Loesche, W. J. (1980) Modulation of immunoreactivity to periodontal disease-associated microorganisms during pregnancy. *Infection and Immunity* **28**, 713–718.
- Luo, G., Abrahams, V. M., Tadesse, S., Funai, E. F., Hodgson, E. J., Gao, J. & Norwitz, E. R. (2010) Progesterone inhibits basal and TNF-alpha-induced apoptosis in fetal membranes: a novel mechanism to explain progesterone-mediated prevention of preterm birth. *Reproductive Sciences* **17**, 532–539.
- Machuca, G., Khoshfeiz, O., Lacalle, J. R., Machuca, C. & Bullon, P. (1999) The influence of general health and socio-cultural variables on the periodontal condition of pregnant women. *Journal of Periodontology* **70**, 779–785.
- Mariotti, A. (1994) Sex steroid hormones and cell dynamics in the periodontium. *Critical Reviews in Oral Biology and Medicine* **5**, 27–53.
- Mariotti, A. (1999) Dental plaque induced gingival diseases. *Annals of Periodontology* **4**, 7–17.
- Marsh, P. D. & Devine, D. A. (2011) How is the development of dental biofilms influenced by the host? *Journal of Clinical Periodontology* **38**(Suppl. 11), 28–35.
- Mascarenhas, P., Gapski, R., Al-Shammari, K. & Wang, H. L. (2003) Influence of sex hormones on the periodontium. *Journal of Clinical Periodontology* **30**, 671–681.
- Mellor, A. L. & Munn, D. H. (2000) Immunology at the maternal-fetal interface: lessons for T cell tolerance and suppression. *Annual Review of Immunology* **18**, 367–391.
- Meulenberg, P. M. & Hofman, J. A. (1989) Salivary progesterone excellently reflects free and total progesterone in plasma during pregnancy. *Clinical Chemistry* **35**, 168–172.
- Michalowicz, B. S., Novak, M. J., Hodges, J. S., DiAngelis, A., Buchanan, W., Papapanou, P. N., Mitchell, D. A., Ferguson, J. E., Lupo, V., Bofill, J., Matseoane, S., Steffen, M. & Ebersole, J. L. (2009) Serum inflammatory mediators in pregnancy: changes after periodontal treatment and association with pregnancy outcomes. *Journal of Periodontology* **80**, 1731–1741.
- Miyazaki, H., Yamashita, Y., Shirahama, R., Goto-Kimura, K., Shimada, N., Sogame, A. & Takehara, T. (1991) Periodontal condition of pregnant women assessed by CPITN. *Journal of Clinical Periodontology* **18**, 751–754.
- Mombelli, A., Schmid, B., Rutar, A. & Lang, N. P. (2000) Persistence patterns of *Porphyromonas gingivalis*, *Prevotella intermedia/nigrescens*, and *Actinobacillus actinomycetemcomitans* after mechanical therapy of periodontal disease. *Journal of Periodontology* **71**, 14–21.
- Morishita, M., Aoyama, H., Tokumoto, K. & Iwamoto, Y. (1988) The concentration of salivary steroid hormones and the prevalence of gingivitis at puberty. *Advances in Dental Research* **2**, 397–400.
- Muramatsu, Y. & Takaesu, Y. (1994) Oral health status related to subgingival bacterial flora and

- sex hormones in saliva during pregnancy. *The Bulletin of Tokyo Dental College* **35**, 139–151.
- Offenbacher, S., Barros, S. P., Singer, R. E., Moss, K., Williams, R. C. & Beck, J. D. (2007) Periodontal disease at the biofilm-gingival interface. *Journal of Periodontology* **78**, 1911–1925.
- Offenbacher, S., Lin, D., Strauss, R., McKaig, R., Irving, J., Barros, S. P., Moss, K., Barrow, D. A., Hefli, A. & Beck, J. D. (2006) Effects of periodontal therapy during pregnancy on periodontal status, biologic parameters, and pregnancy outcomes: a pilot study. *Journal of Periodontology* **77**, 2011–2024.
- O'Neil, T. C. (1979a) Maternal T-lymphocyte response and gingivitis in pregnancy. *Journal of Periodontology* **50**, 178–184.
- O'Neil, T. C. (1979b) Plasma female sex-hormone levels and gingivitis in pregnancy. *Journal of Periodontology* **50**, 279–282.
- Piccinni, M. P. (2010) T cell tolerance towards the fetal allograft. *Journal of Reproductive Immunology* **85**, 71–75.
- Raber-Durlacher, J. E., van Steenberg, T. J., Van der Velden, U., de Graaff, J. & Abraham-Inpijn, L. (1994) Experimental gingivitis during pregnancy and post-partum: clinical, endocrinological, and microbiological aspects. *Journal of Clinical Periodontology* **21**, 549–558.
- Raber-Durlacher, J. E., Zeijlemaker, W. P., Meinesz, A. A. & Abraham-Inpijn, L. (1991) CD4 to CD8 ratio and in vitro lymphoproliferative responses during experimental gingivitis in pregnancy and post-partum. *Journal of Periodontology* **62**, 663–667.
- Samant, A., Malik, C. P., Chabra, S. K. & Devi, P. K. (1976) Gingivitis and periodontal disease in pregnancy. *Journal of Periodontology* **47**, 415–418.
- Sanz, M. & van Winkelhoff, A. J. (2011) Periodontal infections: understanding the complexity-Consensus of the Seventh European Workshop on Periodontology. *Journal of Clinical Periodontology* **38**(Suppl. 11), 3–6.
- Sanz, M., van Winkelhoff, A. J., Herrera, D., Delleijm-Kippuw, N., Simón, R. & Winkel, E. (2000) Differences in the composition of the subgingival microbiota of two periodontitis populations of different geographical origin. A comparison between Spain and The Netherlands. *European Journal of Oral Sciences* **108**, 383–392.
- Shiau, H. J. & Reynolds, M. A. (2010) Sex differences in destructive periodontal disease: exploring the biologic basis. *Journal of Periodontology* **81**, 1505–1517.
- Silness, J. & Løe, H. (1964) Periodontal disease in pregnancy. II. Correlation between oral hygiene and periodontal condition. *Acta Odontologica Scandinavica* **22**, 121–135.
- Stroup, D. F., Berlin, J. A., Morton, S. C., Olkin, I., Williamson, G. D., Rennie, D., Moher, D., Becker, B. J., Sipe, T. A. & Thacker, S. B. (2000) Meta-analysis of observational studies in epidemiology: a proposal for reporting. Meta-analysis of observational studies in epidemiology (MOOSE) group. *The Journal of the American Medical Association* **283**, 2008–2012.
- Syed, S. A. & Loesche, W. J. (1972) Survival of human dental plaque flora in various transport media. *Applied Microbiology* **24**, 638–644.
- Szeker-Bartho, J. (2002) Immunological relationship between the mother and the fetus. *International Reviews of Immunology* **21**, 471–495.
- Takeuchi, Y., Umeda, M., Sakamoto, M., Benno, Y., Huang, Y. & Ishikawa, I. (2001) *Treponema socranskii*, *Treponema denticola*, and *Porphyromonas gingivalis* are associated with severity of periodontal tissue destruction. *Journal of Periodontology* **72**, 1354–1363.
- Telleria, C. M., Ou, J., Sugino, N., Ferguson, S. & Gibori, G. (1998) The expression of interleukin-6 in the pregnant rat corpus luteum and its regulation by progesterone and glucocorticoid. *Endocrinology* **139**, 3597–3605.
- Tilakaratne, A., Soory, M., Ranasinghe, A. W., Corea, S. M., Ekanayake, S. L. & de Silva, M. (2000) Periodontal disease status during pregnancy and 3 months post-partum, in a rural population of Sri-Lankan women. *Journal of Clinical Periodontology* **27**, 787–792.
- Wegmann, T. G., Lin, H., Guilbert, L. & Mosmann, T. R. (1993) Bidirectional cytokine interactions in the maternal-fetal relationship: is successful pregnancy a TH2 phenomenon? *Immunology Today* **14**, 353–356.
- Weitzmann, M. N. & Pacifici, R. (2006) Estrogen regulation of immune cell bone interactions. *Annals of the New York Academy of Sciences* **1068**, 256–274.
- van Winkelhoff, A. J., Loos, B. G., van der Reijden, W. A. & van der Velden, U. (2002) *Porphyromonas gingivalis*, *Bacteroides forsythus* and other putative periodontal pathogens in subjects with and without periodontal destruction. *Journal of Clinical Periodontology* **29**, 1023–1028.
- Yalcin, F., Eskinazi, E., Soyuncu, M., Basesmez, C., Issever, H., Isik, G., Berber, L., Has, R., Sabuncu, H. & Onan, U. (2002) The effect of sociocultural status on periodontal conditions in pregnancy. *Journal of Periodontology* **73**, 178–182.
- Yang, H. W., Huang, Y. F. & Chou, M. Y. (2004) Occurrence of *Porphyromonas gingivalis* and *Tannerella forsythensis* in periodontally diseased and healthy subjects. *Journal of Periodontology* **75**, 1077–1083.
- Yokoyama, M., Hinode, D., Matsuda, K., Yoshioka, M. & Grenier, D. (2005) Effect of female sex hormones on *Campylobacter rectus* and human gingival fibroblasts. *Oral Microbiology and Immunology* **20**, 239–243.
- Zaki, K., el Hak, R., Amer, W., Saleh, F., El Faras, A., Ragab, L. & Nour, H. (1984) Salivary female sex hormone levels and gingivitis in pregnancy. *Biomedica Biochimica Acta* **43**, 749–754.
- Zhao, L., Wu, Y. F., Meng, S., Yang, H., OuYang, Y. L. & Zhou, X. D. (2007) Prevalence of fimA genotypes of *Porphyromonas gingivalis* and periodontal health status in Chinese adults. *Journal of Periodontal Research* **42**, 511–517.

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Clinical relevance

Scientific rationale for study: Different aetiological pathways have been proposed in an attempt to explain the increased gingival inflammation observed during pregnancy, but the results have been inconclusive. Simultaneous

evaluation of the different potential mechanisms may provide a more thorough understanding of this gingival endocrinopathy.

Principal findings: Gingival inflammation during pregnancy was modulated by quantitative and qualitative

variations of the supra- and subgingival biofilm.

Practical implications: These findings suggest that optimal plaque control may be particularly beneficial during pregnancy to control the severity of gingival inflammation.