

## RESEARCH LETTER

# The influence of hydrogen sulfide on gingival wound healing: An in vitro study

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## 1 | INTRODUCTION

Halitosis is a highly prevalent condition, affecting roughly 30% of the world population.<sup>1</sup> Approximately 80% of halitosis cases has an intra-oral origin, mostly due to factors like tongue coating or periodontal diseases.<sup>2</sup>

Halitosis is caused by the release of volatile sulfur compounds (VSC), being hydrogen sulfide ( $H_2S$ ) the most often found VSC in oral cavity.<sup>3</sup> Recent studies have shown deleterious effects of high concentrations of  $H_2S$  to periodontal tissues, and more specifically to gingival epithelial cells (GECs).<sup>4</sup>

Wound healing processes are of paramount importance for the achievement of favorable treatment outcomes of periodontitis and peri-implantitis,<sup>5</sup> and GECs are known to play a key role in this process.

The prevalence of halitosis on periodontal patients is high.<sup>6</sup> Therefore, the aim of this in vitro study is to investigate the effects of different clinically relevant concentrations of  $H_2S$  on GECs. We hypothesize that the presence of  $H_2S$  at concentrations observed in intra-oral halitosis patients will impair the migration, viability, and function of GECs.

## 2 | METHODS

GECs of a human gingival epithelial line (Ca9-22) were plated in a migration insert (migration-insert2 Well24, ibiTreat, Martinsried, Germany) to create a fixed gap between GECs fronts after its retrieval. Therefore, this device allowed to mimic a wound.

$H_2S$  was produced in a Kipp's apparatus and subsequently transferred to the GECs in a container. GECs were incubated for 8 h with various ranges of clinically relevant concentrations of  $H_2S$  (control group: 0 ppb, low: 90–299 ppb, medium: 300–999 ppb, and high:  $\geq 1000$  ppb).<sup>7,8</sup>

$H_2S$  concentration (ppb) inside each container was determined with a gas chromatograph device OralChroma2, at two time points: 15 min after  $H_2S$  insufflation ( $T_0$ ) and after 8-h incubation ( $T_1$ ) to ensure the  $H_2S$  level in the container.

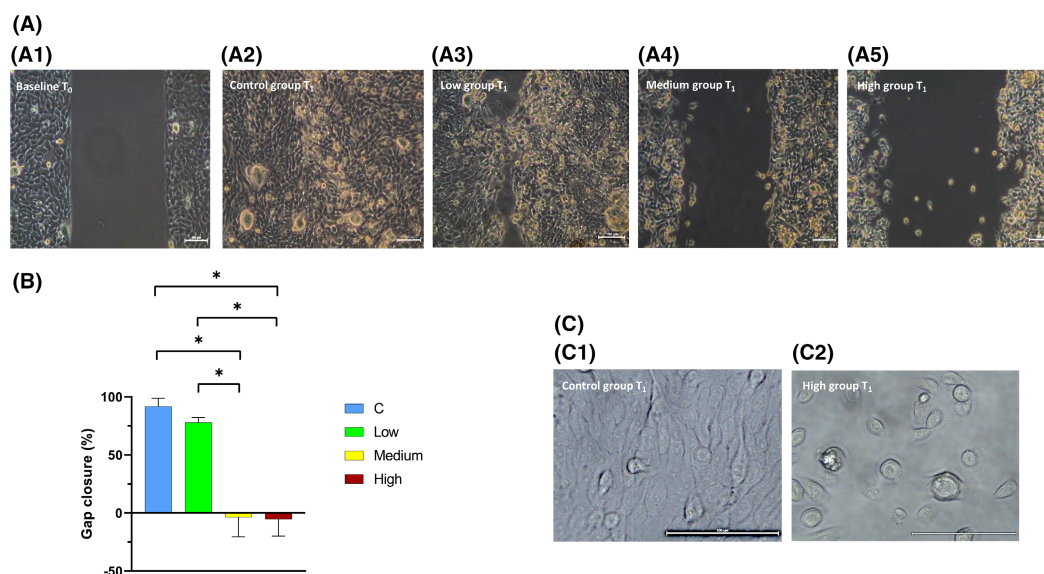
Microscopy analyses of the gap were performed at  $T_0$  and  $T_1$  by using a converted microscope at 100 $\times$  and 400 $\times$  magnification. Subsequently, the cells were trypsinized and prepared for further assessment of cell viability and qPCR analysis for ICAM-1, *e-cadherin*, *TNF- $\alpha$* , *COX IV*, *Ki67*, and *caspase 3*.

The study was approved by the Local ACTA Ethical Committee (020.277).

## 3 | RESULTS

### 3.1 | Less GEC migration with increasing $H_2S$ concentration

A significant difference in gap closure was found between the control and medium  $H_2S$  groups (mean  $\Delta 95.37\%$ ,  $p < 0.0001$ ) and between the control and high  $H_2S$  groups (mean  $\Delta 97.12\%$ ,  $p < .0001$ ) (Figure 1A,B). Similarly, the low  $H_2S$  group showed significant differences in gap closure compared to the medium (mean  $\Delta 81.63\%$ ,



**FIGURE 1** (A) Micrographs of the epithelial cell gap closure: (A1) The baseline (T<sub>0</sub>), (A2) Control group (0 ppb H<sub>2</sub>S) shows a complete closure of the gap after 8-h incubation (T<sub>1</sub> mean 91.64% ± 7.17), (A3) Low concentration group (90–299 ppb H<sub>2</sub>S) shows almost a complete closure of the gap after 8-h incubation (T<sub>1</sub> mean 77.90% ± 4.38), (A4) Medium concentration group (300–999 ppb H<sub>2</sub>S) shows minimum-to-no closure of the gap after 8-h incubation (T<sub>1</sub> mean 3.73% ± 16.70), (A5) High concentration group (≥1000 ppb H<sub>2</sub>S) shows no closure of the gap and even a widening of the gap surface area after 8-h incubation (T<sub>1</sub> mean closure -5.48% ± 14.43). Magnification ×100, Bar = 100 μm. (B) Between-group comparison of the gingival epithelial cell gap closure (T<sub>0</sub>–T<sub>1</sub>). Mean ± SD. \**p* < .0001 C, Control group (0 ppb H<sub>2</sub>S), Low: Low H<sub>2</sub>S concentration group (90–299 ppb H<sub>2</sub>S), Medium: Medium H<sub>2</sub>S concentration group (300–999 ppb H<sub>2</sub>S), High: High H<sub>2</sub>S concentration group (≥1000 ppb H<sub>2</sub>S). (C) Micrographs of gingival epithelial cells (GECs) after 8-h exposure (T<sub>1</sub>) to H<sub>2</sub>S: (C1) GECs from the Control group (0 ppb H<sub>2</sub>S) at T<sub>1</sub>. (C2) GECs from High H<sub>2</sub>S concentration group (≥1000 ppb H<sub>2</sub>S) at T<sub>1</sub>. Magnification × 400. Bar = 100 μm.

*p* < .0001) and high groups (mean Δ83.38%, *p* < .0001). All other comparisons were nonsignificant.

### 3.2 | Less viability and changed morphology of GECs at high H<sub>2</sub>S concentration

Viability of GECs in the medium and high H<sub>2</sub>S groups decreased significantly compared to the control group (mean Δ4.97%, *p* = .0300 and 5.32%, *p* = .0193, respectively). No other significant differences in viability were observed between groups.

Morphometric analysis disclosed a round morphology along with broad intercellular spaces for GECs exposed to high H<sub>2</sub>S concentrations. In contrast, GECs from the control group displayed an elongated morphology, with tight intercellular spaces (Figure 1C).

### 3.3 | E-cadherin, TNF-α, ICAM-1, and Ki-67 are downregulated, whereas caspase 3 is upregulated by high H<sub>2</sub>S concentration

Gene expression of *e-cadherin* (*p* = .009) and *TNF-α* (*p* = .043) were significantly downregulated in the high H<sub>2</sub>S group (Figure 2, Figure S2). Gene expression of *ICAM-1* and *Ki-67* was significantly downregulated, while *caspase 3* significantly upregulated in medium (*p* = .099, *p* = .015 and *p* = .010, respectively) and high (*p* = .099, *p* = .034, and *p* = .015, respectively) H<sub>2</sub>S groups. All other comparisons in gene expression were not statistically significant.

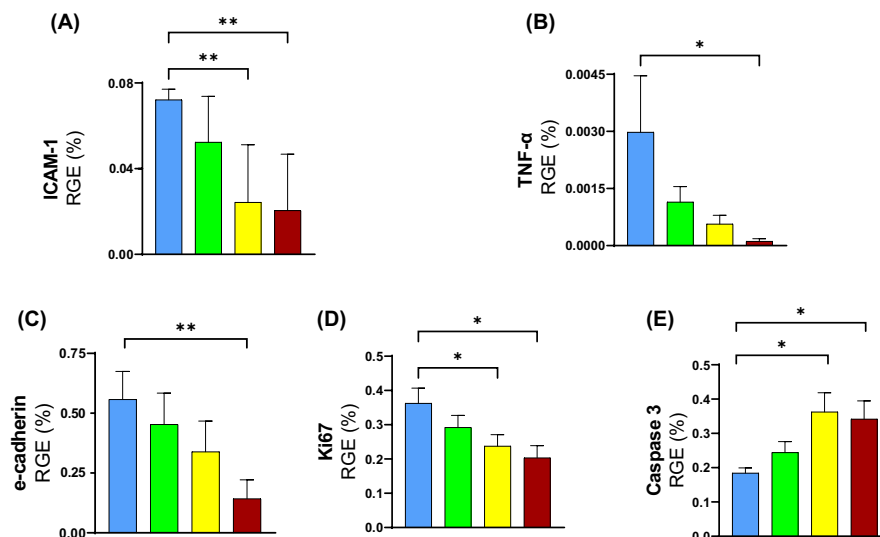
## 4 | DISCUSSION

The current study indicates that H<sub>2</sub>S concentrations ≥300 ppb (Figure S1) may impair gingival wound healing due to (1) impaired gingival epithelial cell migration; (2) reduced cell viability, which may lead to inadequate tissue regeneration and repair; (3) downregulation of genes involved in cellular functions, such as *TNF-α* (inflammation), *e-cadherin* (intercellular junction), *ICAM-1* (cellular adhesion), and *Ki-67* (nuclear function); and (4) upregulation of apoptotic genes (*caspase-3*) leading to an increase in cellular damage to GEC.<sup>9</sup> Furthermore, previous studies have demonstrated that high concentrations of H<sub>2</sub>S can also disrupt the intestinal epithelial layer by decreasing mitochondrial energy metabolism.<sup>10</sup>

There are several limitations given the current study. First, the use of gingival fibroblasts in addition to GECs could increase the understanding of gingival wound healing. Also, the in vitro nature of this study may prevent drawing solid conclusions at clinical level. For example, GECs were continuously exposed to only H<sub>2</sub>S, while in vivo H<sub>2</sub>S concentrations in the oral cavity fluctuate. Furthermore, other VSCs such as methyl mercaptan and dimethyl sulfide can be found in the oral cavity.

Based on the results of this study, we speculate that early diagnosis and treatment of halitosis may play a role in the treatment and prevention of periodontal and peri-implant diseases. Although speculative, it underscores the potential importance of addressing halitosis as part of comprehensive periodontal and peri-implant care. Early halitosis intervention, for example through tongue cleaning and anti-halitosis mouthwashes, could potentially improve outcomes in these patients.

**FIGURE 2** Between-group comparison of relative gene expression (%) of (A) ICAM-1, (B) TNF- $\alpha$ , (C) *e*-cadherin, (D) Ki-67, and (E) caspase 3 at T<sub>1</sub>: Mean  $\pm$  SD are shown. \* $p$  < .05, \*\* $p$  < .01, NS, No statistically significant difference. C, Control group (0 ppb H<sub>2</sub>S), Low: H<sub>2</sub>S concentration 90–299 ppb, Medium: H<sub>2</sub>S concentration 300–999 ppb, High: H<sub>2</sub>S concentration  $\geq$ 1000 ppb.



## AUTHOR CONTRIBUTIONS

J. Villalba-Recuerda contributed to the conception, design, data acquisition, analysis and interpretation, drafted and critically revised the manuscript. I.D.C. Jansen contributed to conception, design, data analysis and interpretation and critically revised the manuscript. M.L. Laine contributed to conception, design, data analysis and interpretation and critically revised the manuscript. All authors gave their final approval and agree to be accountable for all aspects of the work.

## ACKNOWLEDGEMENTS

The authors would like to acknowledge Hadiel Aldarij, Ton Schoenmaker, and Cor Semeins for their contribution.

## FUNDING INFORMATION

The authors do not have any financial interests, either directly or indirectly, in the products or information presented in the paper. The study was entirely financed by the Department of Periodontology, Academic Centre for Dentistry Amsterdam (ACTA), Amsterdam, the Netherlands.

## CONFLICT OF INTEREST STATEMENT

No conflicts of Interest to disclose.

## DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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## SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

**How to cite this article:** Villalba-Recuerda J, Jansen IDC, Laine ML. The influence of hydrogen sulfide on gingival wound healing: An in vitro study. *J Periodont Res*. 2024;00:1-3. doi:[10.1111/jre.13339](https://doi.org/10.1111/jre.13339)